Fundamental and Applied Agriculture

Vol. 6(4), pp. 444[–459:](#page-15-0) 2021

[doi: 10.5455/faa.46026](http://dx.doi.org/10.5455/faa.46026)

AGRONOMY | ORIGINAL ARTICLE

Competence of different priming agents for increasing seed germination, seedling growth and vigor of wheat

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Cite this article: Mim TF, Anwar MP, Ahmed M, Sriti N, Moni EH, Hasan AK, Yeasmin S. 2021. Competence of different priming agents for increasing seed germination, seedling growth and vigor of wheat. Fundamental and Applied Agriculture 6(4): 444[–459.](#page-15-0) [doi: 10.5455/faa.46026](http://dx.doi.org/10.5455/faa.46026)

1 Introduction

Wheat (*Triticum aestivum* L.) is an important crop considering first in terms of area and third in terms of production out of three major cereal crops i.e. maize, rice and wheat in the world [\(Gahtyari et al.,](#page-13-0) [2017\)](#page-13-0). Wheat grain comprises three groups of major components namely starch, protein, and cell wall polysaccharides (dietary fiber), and a range of minor components beneficial to human health [\(Shewry et al.,](#page-15-1) [2013\)](#page-15-1). It is consumed as the major staple food worldwide and has been the most abundant source of calories and protein in the human diet alongside supplying nearly 20% of the total dietary protein [\(Braun et al.,](#page-12-0) [2010\)](#page-12-0). Global wheat production is expected to reach a new record of 780 million tons in 2021, according to a preliminary forecast [\(FAOSTAT,](#page-13-1) [2021\)](#page-13-1). Wheat is considered the second most important cereal crop in Bangladesh after rice [\(Islam,](#page-13-2) [2021\)](#page-13-2). A unique feature of wheat in Bangladesh is the 100% adoption of modern high yielding varieties [\(Rahman and Hasan,](#page-15-2) [2009\)](#page-15-2).

Wheat is now becoming an indispensable food item of the people of Bangladesh. Day by day its demand is raising due to changes in lifestyle and food habits [\(Barma et al.,](#page-12-1) [2019\)](#page-12-1). The total area under the wheat crop of FY 2019-20 has been estimated as 3,32,274 hectares compared to 3,30,348 hectares of FY 2018-19 [\(BBS,](#page-12-2) [2020\)](#page-12-2), 0.58% higher than the previous year. Total production of wheat in FY 2019-20, has been estimated as 10,29,354 metric tons compared to 10,16,811 metric tons in the FY 2018-19, which is 1.23% higher [\(BBS,](#page-12-2) [2020\)](#page-12-2). Despite that Bangladesh has been emerged as the fifth biggest importer of wheat in recent years [\(FAOSTAT,](#page-13-3) [2019\)](#page-13-3). However, sustainable wheat production in Bangladesh is threatened by several stresses (biotic and abiotic) in addition to the competition with other winter crops [\(Barma et al.,](#page-12-1) [2019\)](#page-12-1). According to [Zampieri et al.](#page-15-3) [\(2017\)](#page-15-3), among the abiotic stresses heat stress is most severe to affect crop growth and productivity under the context of global climate change. An estimated 6% fall in wheat production occurs globally for every 1 °C rise in temperature [\(Asseng et al.,](#page-12-3) [2014\)](#page-12-3). Due to increasing temperature, it was estimated that Bangladesh would be 1 °C warmer by 2020 compared to 1971 [\(Kamrul](#page-14-0) [and Hore,](#page-14-0) [2021\)](#page-14-0).

Seed priming, partial hydration of seeds without radicle emergence [\(Farooq et al.,](#page-13-4) [2007b\)](#page-13-4), is a technique that improves the germination, seedling emergence, growth, and yield attributes of a crop [\(Pant and Bose,](#page-14-1) [2016\)](#page-14-1). It is a controlled hydration process that is followed by redrying and triggers many of the physiological processes associated with the early phase of germination and prepares the seed for radicle protrusion which suspends the seeds in the lag phase [\(Paparella et al.,](#page-14-2) [2015\)](#page-14-2). Acceleration of germination in prime seeds can be due to the increasing activity of the degrading enzymes, such as α -amylase, synthesis

of RNA and DNA, the amount of ATP, and the number of mitochondria [\(Afzal et al.,](#page-11-0) [2002\)](#page-11-0). Seed priming can break seed dormancy, curtail seedling emergence time, improve seedling vigor, and leads to better germination and growth of plants [\(Mondal et al.,](#page-14-3) [2011;](#page-14-3) [Srivastava and Bose,](#page-15-4) [2012\)](#page-15-4). Primed seeds in solutions of macro and micronutrients have been shown to improve germination and seedling vigor of wheat [\(Joshi](#page-14-4) [et al.,](#page-14-4) [2018;](#page-14-4) [Hussain et al.,](#page-13-5) [2019;](#page-13-5) [Farooq et al.,](#page-13-6) [2020a;](#page-13-6) [Rai-Kalal and Jajoo,](#page-15-5) [2021\)](#page-15-5), rice [\(Mamun et al.,](#page-14-5) [2018;](#page-14-5) [Anwar et al.,](#page-12-4) [2021\)](#page-12-4), maize [\(Arief et al.,](#page-12-5) [2020\)](#page-12-5) and other crops (Prażak et al., [2020;](#page-15-6) [Farooq et al.,](#page-13-7) [2020b\)](#page-13-7).

Generally, various seed priming techniques, which include hydropriming, osmopriming, chemical priming, nutrient priming, hormonal priming, and redox priming, are used to induce pre-germination changes [\(Paparella et al.,](#page-14-2) [2015;](#page-14-2) [Hussain et al.,](#page-13-5) [2019\)](#page-13-5). Seed pretreatment, either by coating [\(Wei et al.,](#page-15-7) [2014\)](#page-15-7) or pre-soaking (polyamines) [\(Chunthaburee](#page-13-8) [et al.,](#page-13-8) [2014\)](#page-13-8), was found to maintain enhanced tolerance when plants were exposed to stress conditions. Primed seeds showed the higher activities of many enzymes for metabolism of proteins (proteases), carbohydrates (α and β amylases), and lipids (isocitrate lyase), involved in the mobilization of stored reserves in seed, and these enzymes play a pivot role in the breakdown of the macromolecules for embryo growth and development which exerts a positive influence on early and better seedling emergence [\(Varier et al.,](#page-15-8) [2010;](#page-15-8) [Sisodia et al.,](#page-15-9) [2018\)](#page-15-9). Priming is a double technology to enhance rapid and uniform emergence in many crop species, thus achieve high vigor and better yields [\(Farooq et al.,](#page-13-9) [2007a\)](#page-13-9).

Therefore, seed priming approach should be explored to mitigate high-temperature stress of wheat under late sown condition in Bangladesh. Considering the above facts, the present research work was undertaken to assess the performance of a range of seed priming techniques on seed germination and seedling vigor of wheat and to recognize the best priming technique for increased germination and enhanced seedling vigor of wheat.

2 Materials and Methods

2.1 Experimental site and duration

The laboratory experiment was conducted at the Agro Innovation Laboratory, Department of Agronomy, Bangladesh Agricultural University during the 2nd week of November in 2019.

2.2 Experimental treatments and design

The experiment comprised two factors namely priming agent (23 levels) and priming duration (2 levels) arranged in a Completely Randomized Design (CRD)

with 4 replications. Priming agents included Control (No priming), Hydropriming (distilled water), 10000 ppm NaCl, 20000 ppm NaCl, 30000 ppm NaCl, 50 ppm PEG, 100 ppm PEG, 150 ppm PEG, 1 ppm Na_2MoO_4 , 2 ppm Na_2MoO_4 , 3 ppm Na_2MoO_4 , 5000 ppm ZnSO⁴ , 10000 ppm ZnSO⁴ , 15000 ppm ZnSO⁴ , 25 ppm CuSO $_4$, 50 ppm CuSO $_4$, 75 ppm CuSO $_4$, 10000 ppm KCl, 20000 ppm KCl, 30000 ppm KCl, 10000 ppm CaCl $_2$, 20000 ppm CaCl $_2$ and 30000 ppm CaCl $_2$. Priming durations were 6 and 12 hrs.

2.3 Plant material used

BARI Gom-33, developed by Bangladesh Agricultural Research Institute (BARI) collaborated with the International Maize and Wheat Improvement Center (CIMMYT) in 2017, was used as the plant material in the present study. It was a simple cross between Kachu and Solala. Kachu is a 'Kauz'-derived highyielding wheat variety carrying a 2NS segment for blast resistance and Solala is derived from a CIM-MYT's pre-breeding cross, involving a durum wheat derivative that provides a 5–8% yield benefit and is 50-55 ppm zinc (Zn) enriched. It was found with 1.8% disease severity comparing with other varieties. BARI Gom-33 is grown in rabi (winter) season in Bangladesh. The optimum sowing time is between 15 November and 30 November. Generally, 110-115 days are required from sowing to harvest. The yield is 3.95 t ha $^{-1}$. It is high yielding, moderately drought tolerant, grain white glossy and medium in size. This blast-resistant variety is also resistant to leaf rust disease and tolerant to leaf spot disease [\(BARI,](#page-12-6) [2007\)](#page-12-6).

2.4 Priming agents used

All the priming agents used in the experiment were of laboratory grade. Details of the priming agents are presented in [Table 1.](#page-3-0)

2.5 Seed priming

Seeds were soaked in different priming agent solution (previously prepared using distilled water) as per treatments for 6 hours or 12 hours at room temperature (25 \pm 2 °C). The ratio of seed weight to solution volume was 1:5 (g L⁻¹). Then, seeds were removed from the priming agent and, wiped lightly with blotting paper, and allowed to dry by forced air. Dried seeds were put in brown envelopes with tags in sealed polythene bags and stored in a refrigerator at 5 ± 1 °C until being placed for germination. While control treatment received no prior seed priming.

2.6 Germination medium and seed placement

Sterilized sand was used as germination media and plastic-made Petri dishes of 90 mm diameter with 15 mm depth were used as containers. The moisture content of the media was maintained at around 80% of field capacity by watering with distilled water as necessary. One hundred seeds were placed manually on moist soil maintaining more or less equal distance in each petri dish. Petri dishes were put on the desk of the laboratory at room temperature 25 ± 2 °C. Every petri dish was monitored regularly to maintain proper moisture levels. No occurrence of insects or diseases was recorded.

2.7 Observations made

Data were collected on germination percentage, mean germination time, germination index, seedling vigor index, germination co-efficient, seedling root and shoot length, root-shoot ratio, root, shoot and seedling dry weight.

2.8 Germination percentage (GP)

The number of germinated seeds was counted on the 7th day. The appearance of plumule over the sand layer was considered germination.

$$
GP = \frac{S_G}{S_T} \times 100\tag{1}
$$

where, $GP =$ germination (%), S_G and S_T denote the numbers of seed germinated and total number of seeds set for germination, respectively.

2.9 Mean germination time (MGT)

$$
MGT = \frac{\sum D_n}{n}
$$
 (2)

where *n* is the number of seeds germinated on day *D*, and *D* is the number of the day counted from the beginning of germination.

Germination index (GI)

$$
GI = \frac{G_i}{D_i} + \ldots + \frac{G_n}{D_n} \tag{3}
$$

where G_i = number of seeds germinated on *i*th day (D_i) , and G_n = number of seeds germinated on final counting day (*Dn*).

2.10 Seedling vigor index (SVI)

After seed placement for germination, on the 7th day, 5 seedlings from each replicate were randomly selected. Root and shoot lengths were measured, and then oven-dried at 70 °C for 72 hrs. to record the root

Sl. no.	Priming agent	Chemical formula	Manufacturer
	Sodium chloride	NaCl	MERCK, India
	Potassium chloride	KCl.	MERCK, India
3	Calcium chloride	CaCl ₂	MERCK, India
$\overline{4}$	Copper sulfate	CuSO ₄ .5H ₂ O	MERCK, India
	Zinc sulfate	ZnSO ₄ .7H ₂ O	MERCK, India
6	Sodium molybdate	$Na2MoO4.2H2O$	MERCK, India
	Polyethylene glycol 4000	PEG 4000	LOBAL Chemie, India

Table 1. Description of the priming agents

and shoot dry weight of seedlings. Seedling vigor index (SVI) was calculated as follows:

$$
SVI = \frac{L \times GP}{100} \tag{4}
$$

where $L =$ seedling length (root length + shoot length) in cm, and *GP* = germination percentage.

2.11 Germination co-efficient (GC)

The co-efficient of germination was calculated using the following formula (Copeland 1976).

$$
GC = \frac{N100 (A_1 + A_2 + \dots + A_n)}{A_1 T_1 + A_2 T_2 + \dots + A_n T_n}
$$
(5)

Where *A* is the number of seeds germinated, *T* is the time corresponding to A , $n =$ Number of days to the final count.

2.12 Seedling growth

On the 7th day of seed placement for germination, 5 seedlings from each replicate were randomly selected. Root length was measured from the base of the plant up to the end of the longest root and expressed in mm. Shoot length was measured from the base of the plant up to the tip of the longest leaf and expressed in mm. The root-shoot ratio was calculated as the ratio of root length to shoot length. Root and shoot dry weight of sample seedlings were measured after drying the roots in an oven at 70 \degree C for 72 hrs. Finally, the root and shoot dry weight of each seedling were calculated and expressed in mg.

2.13 Statistical analysis

The recorded data were compiled and tabulated for statistical analysis. Analysis of variance (ANOVA) was done with the help of the computer package MSTAT-C (Statistical software). The mean differences among the treatments were adjudged by Duncan's Multiple Range Test [\(Gomez and Gomez,](#page-13-10) [1984\)](#page-13-10) at 5% level of significance.

3 Results

3.1 Germination and seedling vigor

Priming agent brings a significant effect (p<0.01) on seed germination rate and seedling vigor inof BARI Gom-33 [\(Table 2\)](#page-4-0). In general, seed priming showed a positive effect on seed germination and seedling vigor of wheat. Hydropriming and $CuSO₄$ priming at any concentration reduced germination rate while $CaCl₂$ priming enhanced germination rate the most. Compared to control, seed priming with 10000 ppm $CaCl₂$ took the least mean germination time (1.01 days), and priming with 15000 ppm $\rm ZnSO_4$ resulted in the highest mean germination time (2.95 days). Priming with 10000 ppm CaCl₂, 10000 ppm KCl, 20000 ppm KCl, and 20000 ppm $CaCl₂$ produce the best germination index; on the other hand, priming with $\rm CaCl_2$, 50 ppm PEG, and 100 ppm PEG resulted in the most vigorous seedlings. Duration of priming had a significant effect (p<0.01) on germination rate and seedling vigor in BARI Gom-33. The 6-hour priming duration resulted in higher germination rate and seedling vigor compared to 12-hour priming duration [\(Table 3\)](#page-4-0). Interaction between priming agent and duration of priming did not produce any significant effect (p>0.01) on germination rate, mean germination, germination index, or seedling vigor either [\(Table 4\)](#page-5-0).

3.2 Seedling growth

Priming agent exerted a significant effect (p<0.01) on the root length of BARI Gom-33. Seed priming with CaCl₂, 50 ppm PEG and 150 ppm PEG performed the best in terms of root length while $ZnSO₄$ priming performed the worst. It is interesting to note that $ZnSO₄$ priming resulted in even shorter root length compared to control [\(Table 5\)](#page-7-0). The duration of priming and interaction between priming agent and priming duration had no significant effect (p>0.01) on the root length of BARI Gom-33 [\(Table 6](#page-7-0) and [Table 7\)](#page-8-0). A significant positive effect of seed priming was found on shoot length. Priming agent CaCl $_2$, 50 ppm PEG, and 150 ppm PEG performed the best in terms of shoot length, on the other hand $ZnSO₄$ priming resulted in shorter shoot length compared to control [\(Table 5\)](#page-7-0).

Priming agent	Final GP $(\%)$	MGT (days)	GI	SVI	GC
Control	84.75 i	3.550a	30.751	11.92 j	22.88 h
Hydropriming	89.13 fgh	2.675 cd	42.38 hi	14.21 i	24.90 efgh
10000 ppm NaCl	92.38 cde	1.813 fg	67.13 efg	18.06 cde	25.77 def
20000 ppm NaCl	91.25 defgh	2.063 e	66.13 fg	17.43 def	25.65 def
30000 ppm NaCl	88.38h	1.888 ef	65.13 g	15.78 gh	25.56 defg
50 ppm PEG	96.00 ab	1.450 hi	68.75 cdef	21.26 b	26.91 abcde
100 ppm PEG	94.88 bc	1.612 gh	67.75 defg	20.78 b	26.31 cde
150 ppm PEG	92.88 cde	1.650 fgh	65.75 fg	19.22 c	26.39 bcde
1 ppm $Na2MoO4$	91.88 def	2.563 cd	43.75 h	16.55 fgh	23.41 h
2 ppm Na ₂ MoO ₄	92.50 cde	2.713 bc	41.50 hij	16.73 efg	23.01 h
3 ppm Na ₂ MoO ₄	90.25 efgh	2.600 cd	39.50 ijk	15.28 hi	23.10 h
5000 ppm $ZnSO4$	93.00 cde	2.425 d	38.88 ijk	9.271 k	23.67 fgh
10000 ppm $ZnSO4$	91.63 defg	2.737 bc	37.88 jk	8.545 kl	23.41 gh
15000 ppm $ZnSO4$	90.38 efgh	2.950 b	36.88 k	7.4181	23.38 gh
25 ppm CuSO ₄	90.13 efgh	2.475 cd	41.13 hij	11.11j	23.40 gh
50 ppm CuSO ₄	91.00 defgh	2.600 cd	39.50 ijk	10.68j	23.79 fgh
75 ppm CuSO ₄	88.75 gh	2.688 cd	38.13 jk	9.379 k	23.06 h
10000 ppm KCl	96.88 ab	1.263 ijk	73.38 ab	18.91 c	27.73 abcd
20000 ppm KCl	96.63 ab	1.313 ij	72.13 abc	18.25 cd	27.35 abcd
30000 ppm KCl	94.00 bcd	1.250 ijk	70.50 bcde	17.11 defg	27.55 abcd
10000 ppm $CaCl2$	98.88 a	1.013 k	74.25 ait	22.77 a	28.83a
20000 ppm CaCl ₂	96.63 ab	1.112 jk	72.88 ab	21.32 b	28.55 ab
30000 ppm CaCl ₂	96.25 ab	1.188 jk	70.75 abcd	20.74 b	28.45 abc
$S\bar{x}$	0.92	0.08	1.13	0.45	0.69
Sig. level	**	$***$	$***$	$***$	**
CV(%)	2.81	11.24	5.82	8.1	7.65

Table 2. Effect of priming agent on germination and seedling vigor of BARI Gom-33

 $GP =$ germination percentage, MGT = mean germination time, $GI =$ germination index, $SVI =$ seedling vigour index, and GC = germination co-efficient

Table 3. Effect of priming duration on germination and seedling vigor of BARI Gom-33

Priming duration	Final GP $(\%)$	MGT (days)	GI	SVI	GC
6 hrs	93.20 a	2.08	55.33	16.12a	25.37
12 hrs	91.88 b	2.06	54.65	15.42 b	25.33
$S\bar{x}$	0.27	0.02	0.33	0.13	0.2
Sig. level	$***$	NS	NS	$**$	NS
CV(%)	2.81	11.24	5.82	8.1	7.65

 $GP =$ germination percentage, MGT = mean germination time, $GI =$ germination index, $SVI =$ seedling vigour index, and GC = germination co-efficient

Seed priming duration also had a significant effect (p<0.01) on the shoot length of BARI Gom-33, and 6 hrs priming performed better than 12 hrs priming [\(Ta](#page-7-0)[ble 6\)](#page-7-0). Priming agent and duration failed to interact significantly for shoot length of BARI Gom-33 [\(Ta](#page-8-0)[ble 7\)](#page-8-0). Priming agents showed a significant effect on seedling length of BARI Gom-33. Seed priming with 10000 ppm CaCl $_2$, 20000 ppm CaCl $_2$ and 50 ppm PEG resulted in the highest seedling length, while $ZnSO₄$ priming produced the lowest seeding length which

was even lower than that of control [\(Table 5\)](#page-7-0). The duration of priming made a significant effect $(p<0.01)$ on the seedling length of BARI Gom-33, and priming for 6 hrs resulted in higher seedling length than 12 hrs priming [\(Table 6\)](#page-7-0). Like root length and shoot length, seedling length of BARI Gom-33 was also found unaffected by the interaction between seed priming agent and priming duration [\(Table 7\)](#page-8-0).

Priming agent \times priming duration		Final GP $(\%)$	MGT (days)	GI	$\ensuremath{\mathrm{SVI}}$	GC
Control	6 hrs	84.75	3.55	30.75	11.92	22.88
	12 hrs	84.75	3.55	30.75	11.92	22.88
Hydropriming	6 hrs	89.25	2.85	42.00	14.90	25.13
	12 hrs	89.00	2.50	42.75	13.53	24.68
10000 ppm NaCl	6 hrs	93.75	1.93	68.25	18.38	25.75
	12 hrs	91.00	1.70	66.00	17.73	25.80
20000 ppm NaCl	6 hrs	92.00	2.10	67.25	18.01	25.65
	12 hrs	90.50	2.03	65.00	16.87	25.65
30000 ppm NaCl	6 hrs	89.25	1.78	66.25	16.38	25.50
	12 hrs	87.50	2.00	64.00	15.19	25.63
50 ppm PEG	6 hrs	97.00	1.50	70.25	21.68	27.08
	12 hrs	95.00	1.40	67.25	20.83	26.75
100 ppm PEG	6 hrs	94.75	1.70	68.75	20.89	26.48
	12 hrs	95.00	1.53	66.75	20.66	26.15
150 ppm PEG	6 hrs	93.75	1.70	65.75	19.24	26.53
	12 hrs	92.00	1.60	65.75	19.19	26.25
1 ppm Na ₂ MoO ₄	6 hrs	93.00	2.58	44.00	17.16	23.58
	12 hrs	90.75	2.55	43.5	15.93	23.25
2 ppm $Na2MoO4$	6 hrs	93.00	2.68	40.75	16.91	23.00
	12 hrs	92.00	2.75	42.25	16.54	23.03
3 ppm Na ₂ MoO ₄	6 hrs	91.25	2.50	39.75	15.83	23.15
	$12\,\mathrm{hrs}$	89.25	2.70	39.25	14.73	23.05
5000 ppm $ZnSO4$	6 hrs	93.75	2.40	40.00	9.46	23.75
	12 hrs	92.25	2.45	37.75	9.08	23.60
10000 ppm $ZnSO4$	6 hrs	93.25	2.83	38.75	8.94	23.38
	12 hrs	90.00	2.65	37.00	8.15	23.45
15000 ppm $ZnSO4$	6 hrs	90.5	2.90	36.50	7.64	23.50
	12 hrs	90.25	$3.00\,$	37.25	7.19	23.25
25 ppm CuSO ₄	6 hrs	90.75	2.38	40.75	11.33	22.93
	12 hrs	89.5	2.58	41.50	10.88	23.88
50 ppm CuSO ₄	6 hrs	91.50	2.53	38.75	11.03	23.73
	12 hrs	90.50	2.68	40.25	10.32	23.85
75 ppm CuSO ₄	6 hrs	89.25	2.58	38.25	9.45	23.08
	12 hrs	88.25	2.80	38.00	9.31	23.05
10000 ppm KCl	6 hrs	97.75	1.28	73.75	19.49	27.78
	12 hrs	96.00	1.25	73.00	18.34	27.68
20000 ppm KCl	6 hrs	97.25	1.40	71.5	18.70	27.30
	12 hrs	96.00	1.23	72.75	17.8	27.40
30000 ppm KCl	6 hrs	95.25	1.30	70.75	17.42	27.48
	12 hrs	92.75	1.20	70.25	16.79	27.63
10000 ppm CaCl ₂	6 hrs	99.00	1.10	75.00	23.15	29.08
	12 hrs	98.75	0.93	73.50	22.39	28.58
20000 ppm CaCl ₂	6 hrs	97.25	1.20	73.00	21.69	28.48
	12 hrs	96.00	1.03	72.75	20.95	28.63
30000 ppm CaCl ₂	6 hrs	96.25	1.18	71.75	21.08	28.45
	12 hrs	96.25	1.20	69.75	20.40	28.45
Sî		1.30	0.12	1.60	0.64	0.97
Sig. level		$_{\rm NS}$	$_{\rm NS}$	$_{\rm NS}$	$_{\rm NS}$	NS
CV(%)		2.81	11.24	5.82	8.10	7.65

Table 4. Interaction effects of priming agent and duration on germination and seedling vigor of BARI Gom-33

GP = germination percentage, MGT = mean germination time, GI = germination index, SVI = seedling vigour index, and GC = germination co-efficient, NS = Not significant

3.3 Seedling dry matter

Seed priming agent significantly affected (p<0.01) seedling root dry weight of BARI Gom-33 [\(Table 8\)](#page-9-0). Priming with KCl produced the highest value; on the contrary, no advantages of priming with 75 ppm CuSO $_4$, 15000 ppm ZnSO $_4$, 10000 ppm ZnSO $_4$, and 50 ppm $CuSO₄$ was observed rather they produced statistically lower root dry weight than that of no priming [\(Table 8\)](#page-9-0). Priming duration had no significant effect (p>0.01) on seedling root dry weight of BARI Gom-33 [\(Table 9\)](#page-9-0). Also, no significant effect (p>0.01) was found for the interaction between priming agent and duration on root dry weight of BARI Gom-33 [\(Table 10\)](#page-10-0).

An overall positive significant effect of seed priming was observed on shoot dry weight of BARI Gom-33. Seed priming with 10000 ppm CaCl₂, 20000 ppm $CaCl₂$ and, 50 ppm PEG produced the highest and statistically similar seedling shoot dry weight [\(Ta](#page-9-0)[ble 8\)](#page-9-0). On the contrary, no advantage of 75 ppm CuSO $_4$, 15000 ppm ZnSO $_4$, and 50 ppm CuSO $_4$ priming was observed since they produced statistically lower seedling shoot dry weight than no priming [\(Ta](#page-9-0)[ble 8\)](#page-9-0). Duration of priming also showed a significant effect (p<0.01) on seedling shoot dry weight, and 6 hrs priming was found better than 12 hrs priming [\(Table 8\)](#page-9-0). No significant effect was found for the interaction between priming agent and duration on shoot dry weight of BARI Gom-33 [\(Table 10\)](#page-10-0).

The seedling dry weight of BARI Gom-33 was significantly affected $(p<0.01)$ by seed priming agents. Priming agents like KCl and 10000 ppm CaCl₂ produced the highest seedling dry weight while 75 ppm $CuSO₄$ and 15000 ppm $ZnSO₄$ produced the lowest seedling dry weight which was lower than no priming control [\(Table 8\)](#page-9-0). Priming duration also exerted a significant effect on seedling dry weight, and priming for 6 hrs resulted in higher seedling dry weight than 12 hrs priming [\(Table 9\)](#page-9-0). No significant effect for the interaction between priming agent and duration on the seedling dry weight of BARI Gom-33 was found [\(Table 10\)](#page-10-0). Priming agent, priming duration, and their interaction failed to produce any significant effect on the root-shoot ratio of BARI Gom-33 [\(Tables 8](#page-9-0) to [10\)](#page-10-0).

4 Discussion

Seed germination rate, germination pattern, seedling growth, and vigor are very crucial for plant growth and productivity under both normal and adverse conditions like heat and moisture stresses. Crop plants often pass through a period of abiotic stresses during their life cycle under natural environments which adversely affect their growth and productivity [\(Hussain](#page-13-11) [et al.,](#page-13-11) [2018\)](#page-13-11). As evident from different reports [\(Wahid](#page-15-10) [et al.,](#page-15-10) [2008;](#page-15-10) [Patanè et al.,](#page-14-6) [2009\)](#page-14-6), seed priming provides plants with greater tolerance when exposed to stress.

Therefore, it was, hypothesized that pre-sowing seed treatment could enhance germination, increase germination rate and improve seedling vigor and growth that could be helpful for wheat seedlings to overcome different abiotic stresses under late sown conditions which is now very common under Bangladesh context.

A positive impact of seed priming (except hydropriming and $CuSO₄$ priming) on seed germination percentage of wheat is evident from this study. In general, CaCl₂ priming performed the best. These findings are in harmony with those of many others who confirmed increased and faster germination along with synchronized emergence of primed seeds in wheat and other crop species [\(Khan et al.,](#page-14-7) [2020;](#page-14-7) [Kaczmarek et al.,](#page-14-8) [2016;](#page-14-8) [Anwar et al.,](#page-12-7) [2020\)](#page-12-7). Among the osmopriming agents $CaCl₂$ showed the best activity concerning seed germination, germination speed, and germination index of BRRI dhan40, BRRI dhan41, and BINA dhan7 [\(Islam et al.,](#page-14-9) [2012\)](#page-14-9).

According to [Ajouri et al.](#page-12-8) [\(2004\)](#page-12-8), priming induces a range of biochemical changes in the seed that is required for initiating the germination process i.e., breaking of dormancy, hydrolysis or metabolism of inhibitors, imbibition, and enzymes activation. Priming is an effective strategy to impart abiotic stress tolerance besides strengthening the defense line of crop plants. In seed priming, pre-sowing treatments are applied that control the hydration level within the seed and allow pre-germinative metabolic processes (physiochemical) to proceed while preventing radical emergence [\(Hussain et al.,](#page-13-12) [2015a;](#page-13-12) [Lutts et al.,](#page-14-10) [2016;](#page-14-10) [Zheng et al.,](#page-15-11) [2015\)](#page-15-11).

Plants that emerged from the primed seeds showed vigorous head start and higher stress tolerance mainly because of more effective energy metabolism, OA (Osmotic adjustment), quick cellular defense systems, enlarged embryo, and enhanced enzymatic activation [\(Jisha et al.,](#page-14-11) [2012\)](#page-14-11). Seed priming offers a smart, innovative, realistic, and effective option for achieving faster and uniform emergence, vigorous stand establishment, and higher productivity in crop plants under normal and stressful conditions [\(Hussain et al.,](#page-13-5) [2019\)](#page-13-5). It is also evident from some research work that priming with CuSO_4 treatments on oat (*Avena sativa* L.) seeds had no significant effects on germination speed [\(Iqbal,](#page-13-13) [2020\)](#page-13-13).

This study verified that priming with $CaCl₂$ and PEG promoted seedling length most while $ZnSO₄$ priming inhibited seedling length. It is interesting to note that $ZnSO_4$ priming resulted in a shorter seedling length compared to control. Primed seedlings can produce normal seedlings under stress conditions. [Ashraf and Foolad](#page-12-9) [\(2005\)](#page-12-9) reported that under stress conditions primed seedlings can grow normally without any disturbance. Priming leaves a stress memory in the seed as the pre-germinative soaking process of seeds in the priming technique ex-

Priming agent	Root length (cm)	Shoot length (cm)	Seedling length (cm)
Control	7.200 fg	6.850 g	14.05i
Hydropriming	8.125 f	7.787 f	15.91 h
10000 ppm NaCl	10.69 bcd	8.837 de	19.52 de
20000 ppm NaCl	10.70 bcd	8.375 ef	19.08 ef
30000 ppm NaCl	10.23 cde	7.600 f	17.83 fg
50 ppm PEG	11.68 ab	10.48 ab	22.15 ab
100 ppm PEG	11.63 ab	10.30 ab	21.92 abc
150 ppm PEG	10.86 bc	9.825 bc	20.69 cd
1 ppm Na ₂ MoO ₄	9.688 de	8.338 ef	18.02 fg
2 ppm Na ₂ MoO ₄	9.650 de	8.400 ef	18.05 fg
3 ppm Na ₂ MoO ₄	9.250 e	7.688 f	16.94 gh
5000 ppm $ZnSO4$	5.300 ij	4.675 ij	9.9751
10000 ppm $ZnSO4$	4.838 ij	4.488 jk	9.325 lm
15000 ppm $ZnSO4$	4.412 j	3.787 k	8.200 m
25 ppm CuSO ₄	6.475 gh	5.850h	12.32 j
50 ppm CuSO ₄	6.400 gh	5.338 hi	11.74 jk
75 ppm CuSO ₄	5.600 hi	4.988 ij	10.59 kl
10000 ppm KCl	10.21 cde	9.300 cd	19.51 de
20000 ppm KCl	9.663 de	9.212 cd	18.88 ef
30000 ppm KCl	9.475 e	8.712 de	18.19 efg
10000 ppm $CaCl2$	12.07 a	10.96 a	23.04 a
20000 ppm CaCl ₂	11.49 ab	10.57 ab	22.06 ab
30000 ppm $CaCl2$	11.44 ab	10.11 b	21.55 bc
Sî	0.34	0.26	0.44
Sig. level	$***$	$***$	$***$
CV(%)	10.68	9.39	7.29

Table 5. Effect of different priming agents on seedling growth of BARI Gom-33

 $**$ = Significant at 1% level of probability

** = Significant at 1% level of probability, NS = Not significant

hibits stress exposure that may improve the stress tolerance capacity in seeds during the post-germinative or seedling establishment period [\(Hussain et al.,](#page-13-14) [2015b;](#page-13-14) [Chakraborty and Dwivedi,](#page-12-10) [2021\)](#page-12-10).

Seed priming with $CaCl₂$ performed better in comparison to control. Because $Ca₂⁺$ increases cell-wall integrity and acts as a secondary messenger in signaling pathways of developmental and physiological processes, also $Ca₂⁺$ icon decoded into downstream responses that ultimately lead to defense [\(Thor,](#page-15-12) [2019\)](#page-15-12). The results of this study suggested that the osmopriming of wheat seeds with $CaCl₂$ would be more exact-

ing than other priming agents. Vigorous seedling growth, emergence, and yield performance by $CaCl₂$ priming have also been reported in wheat under late sown conditions [\(Farooq et al.,](#page-13-15) [2008\)](#page-13-15).

[Bose et al.](#page-12-11) [\(2018\)](#page-12-11) stated that seed priming with PEG significantly improved the levels of photosynthetic pigments under abiotic stress conditions. [Lee](#page-14-12) [and Kim](#page-14-12) [\(2000\)](#page-14-12) revealed that priming increased the metabolic activities of seed ultimately gained a substantial shoot length than non-primed seed. Osmopriming with PEG enhanced the ATPase activity in peanuts with substantial improvement in RNA synPriming agent \times priming duration Root length (cm) Shoot length (cm) Seedling length (cm)

S*x*ˆ 0.48 0.37 0.61 Sig. level NS NS NS CV (%) 10.68 9.39 7.29

Table 7. Interaction effect of priming agent and duration on seedling growth of BARI Gom-33

NS = Not significant

Priming agent	Root dry matter (mg)	Shoot dry matter (mg)	Seedling dry matter (mg)	Root: Shoot
Control	3.675 i	3.525k	7.200 i	0.927
Hydropriming	4.463 gh	4.175j	8.637h	0.941
10000 ppm NaCl	4.925 ef	5.275f	10.20 fg	0.818
20000 ppm NaCl	4.863 fg	4.838 gh	9.700 g	0.856
30000 ppm NaCl	4.350 h	4.438 ij	8.788 h	0.856
50 ppm PEG	5.100 def	6.313 abc	11.41 c	0.722
100 ppm PEG	5.025 def	6.125 bcd	11.15 cd	0.73
150 ppm PEG	4.800 fg	5.700 e	10.50 ef	0.738
1 ppm Na ₂ MoO ₄	5.625 с	5.162 fg	10.79 def	0.937
2 ppm Na ₂ MoO ₄	5.537 с	5.012 fg	10.55 ef	0.946
3 ppm Na ₂ MoO ₄	5.325 cde	4.550 hi	9.875 g	1.02
5000 ppm $ZnSO4$	3.388 ij	3.138 lm	6.525 jk	0.972
10000 ppm $ZnSO4$	3.150 jk	3.000 lmn	6.150 jkl	0.916
15000 ppm $ZnSO4$	2.888 kl	2.750 n	5.637 lm	0.923
25 ppm CuSO ₄	3.425 ij	3.1751	6.60j	0.97
50 ppm CuSO ₄	3.200 jk	2.800 mn	6.000 kl	0.986
75 ppm CuSO ₄	2.5631	2.750 n	5.313 m	0.8
10000 ppm KCl	6.863a	6.275 bc	13.14a	0.94
20000 ppm KCl	6.525 ab	6.113 bcd	12.64 ab	0.941
30000 ppm KCl	6.325 b	5.787 de	12.11 b	0.952
10000 ppm CaCl ₂	5.450 cd	6.662a	12.11 b	0.726
20000 ppm CaCl ₂	4.988 ef	6.475 ab	11.46 с	0.705
30000 ppm CaCl ₂	4.950 ef	6.088 cd	11.04 cde	0.721
$S\hat{x}$	0.14	0.12	0.2	0.1
Sig. level	$***$	$***$	**	$_{\rm NS}$
CV(%)	8.66	7.07	5.89	31.16

Table 8. Effect of different priming agents on seedling dry matter of BARI Gom-33

 $**$ = Significant at 1% level of probability, NS = Not significant

** = Significant at 1% level of probability, NS = Not significant

theses and activity of acid phosphatase in the cotyledon and embryonic axis [\(Nawaz et al.,](#page-14-13) [2013\)](#page-14-13). In Arabidopsis seeds, osmopriming (−0.75 MPa PEG 6000) led to the accumulation of α tubulin and β tubulin (*Tubulin subunits*) [\(Kubala et al.,](#page-14-14) [2015\)](#page-14-14). Numerous research demonstrated that osmoprimed seeds are effective to improve germination, emergence, and seedling establishment of several plants, especially under stress conditions like ryegrass, sorghum, tomato, and wheat, Barley [\(Sun et al.,](#page-15-13) [2010;](#page-15-13) [Sale-](#page-15-14) [hzade et al.,](#page-15-14) [2009;](#page-15-14) [Zhang et al.,](#page-15-15) [2015;](#page-15-15) [Amini,](#page-12-12) [2013;](#page-12-12) [Amooaghaie and Nikzad,](#page-12-13) [2013\)](#page-12-13). [Baque et al.](#page-12-14) [\(2016\)](#page-12-14) reported that the highest shoot length of wheat was secured when the seed was primed with PEG. Osmopriming of seeds with PEG has great potential to improve the stand establishment and performance of crop plants under stressful environments [\(Zhang](#page-15-15) [et al.,](#page-15-15) [2015;](#page-15-16) [Salah et al.,](#page-15-16) 2015; Aydinoğlu et al., [2019\)](#page-12-15). Seeds treated with PEG6000 (-0.6 MPa) showed improved germination and healthy seedling growth, im-

Table 10. Interaction effect of different priming agents and duration on seedling dry matter of BARI Gom-33

NS = Not significant

proved root length, and antioxidant defense mechanism [\(Mouradi et al.,](#page-14-15) [2016\)](#page-14-15). The beneficial effects of PEG osmopriming were evident on root growth as reported by [Patanè et al.](#page-14-6) [\(2009\)](#page-14-6).

Priming with KCl produced the highest root dry weight. On the contrary, no advantage of $CuSO₄$ or ZnSO⁴ priming was observed since they produced a statistically lower root dry weight than control. It is because of KCl is a strong stressor that affected dry matter of wheat [\(Natasha,](#page-14-16) [2021\)](#page-14-16). A positive influence of seed priming on seedling growth (length and dry matter) was also reported by [\(Anwar et al.,](#page-12-4) [2021\)](#page-12-4), and KCl and $CaCl₂$ performed the best.

Seed priming with $CaCl₂$ and PEG produced the highest and statistically similar seedling shoot dry weight. On the contrary, $CuSO₄$ and $ZnSO₄$ priming were observed inhibiting the same since they produced lower seedling shoot dry weight than no priming. $\rm CaCl_2$ showed better results in incrementing the shoot length of the Yard-long bean [\(Karim et al.,](#page-14-17) [2020\)](#page-14-17). The highest plumule length of BARI Gom-27 was obtained from seeds pre-treated with 10% PEG solution [\(Baque et al.,](#page-12-14) [2016\)](#page-12-14). Significant positive correlation between root and shoot length, root with root/shoot length ratio, and shoot with root/shoot length ratio were reported by [Baloch et al.](#page-12-16) [\(2012\)](#page-12-16) at 0 and 15% PEG stress levels. Seed priming with PEG (30%) significantly increased the shoot and root length as well as seedling fresh and dry biomass as revealed by [Salah](#page-15-16) [et al.](#page-15-16) [\(2015\)](#page-15-16).

Priming agents KCl and CaCl₂ produced the highest seedling dry weight and $CuSO₄$ and $ZnSO₄$ produced the lowest seedling dry weight. Seed priming with KCl or $CaCl₂$ also improved the seedling growth, stand establishment as well yield performance in direct-seeded rice [\(Farooq et al.,](#page-13-16) [2006\)](#page-13-16). The K^+ is essential for activation of enzymes, for turgor and membrane potential balance, and in osmotic regulation in cells [\(Cherel,](#page-13-17) [2004\)](#page-13-17). Priming improved the K + balance that activates alpha-amylase, a basis for seed invigoration. A similar result was also obtained by [Hasan et al.](#page-13-18) [\(2016\)](#page-13-18) who reported that rice seeds primed with 5% CaCl₂ and 3% KCl for 24 hours produced the highest shoot dry mass of rice seedlings. [Jisha et al.](#page-14-11) [\(2012\)](#page-14-11) reported that the overall growth of plants was enhanced due to the seed-priming treatments. [Maiti et al.](#page-14-18) [\(2013\)](#page-14-18) observed that seed priming increases the seedling vigor of several vegetable crops. Seed germination, radicle/plumule emergence are inhibited by Cu_2^+ at high concentrations and Zn_2^+ at a different dose [\(Ahmed et al.,](#page-12-17) [2021\)](#page-12-17). Previous results demonstrated that seeds primed in 100 ppm CuSO₄ showed a reduction in seedlings shoot length, root length, seedling fresh biomass as compared to $CuSO₄$ at 200 ppm concentration [\(Iqbal,](#page-13-13) [2020\)](#page-13-13).

CaCl₂ and PEG increased seed germination and early seedling growth in two wheat cultivars in both control and drought conditions [\(Asaduzzaman et al.,](#page-12-18) [2021\)](#page-12-18). [Chen et al.](#page-12-19) [\(2021\)](#page-12-19) revealed that seed priming enhanced germination, seedling growth, and water relation behavior of wheat genotypes. All the characters showed the best results when wheat seeds were treated with 10% PEG solution. Osmopriming of rice seeds with KCl and $CaCl₂$ improved the emergence, emergence energy, and seedling emergence index [\(Farooq et al.,](#page-13-16) [2006\)](#page-13-16). Like K^+ , Ca_2^+ also plays very important roles in cell elongation and division, maintains cell wall integrity, regulates the uptake of nutrients across the membrane, and improves uptake of water in plants, and alleviates the adverse effect of Na⁺ during plant growth [\(Patade et al.,](#page-14-19) [2009;](#page-14-19) [Gobi](#page-13-19)[nathan et al.,](#page-13-19) [2009\)](#page-13-19). Germination speed is related to seedling vigor and it could be a significant determinant of good field performance [\(da Cruz and Milach,](#page-13-20) 2004). Priming of barley seeds with CaCl $_2$ improved drought tolerance attributed to enhanced transpiration rate without negative effects on the leaf turgor status and better stomatal aperture [\(Kaczmarek et al.,](#page-14-8) [2016\)](#page-14-8).

Intriguingly, the duration of priming had a significant effect on germination rate and seedling vigor in BARI Gom-33. The 6-hour priming duration showed a higher germination rate and seedling vigor compared to the 12-hour priming duration irrespective of concentration of any priming agent. As reported by [Ibrahim](#page-13-21) [\(2019\)](#page-13-21), priming agent and duration are very important factors that determine the germination success and seedling establishment.

5 Conclusion

In conclusion, present findings confirm the potentiality of seed priming to enhance the seed germination and seedling vigor of wheat, and pre-sowing seed priming with $CaCl₂$ for 6 hours was found the best for BARI Gom-33. Further detailed studies are required to explore the potentiality of seed priming to combat abiotic stresses like heat and moisture stresses in wheat under Bangladesh conditions.

Conflict of Interest

The authors declare that there is no conflict of interests regarding the publication of this paper.

References

Afzal A, Basra MA, Ahmad N, Cheema MA, Warraich EA, Khaliq A. 2002. Effect of priming and growth regulator treatments on emergence and seedling growth of hybrid maize (*Zea mays* L.). International Journal of Agriculture and Biology 4:303–306.

- Ahmed B, Rizvi A, Syed A, Elgorban AM, Khan MS, AL-Shwaiman HA, Musarrat J, Lee J. 2021. Differential responses of maize (*Zea mays*) at the physiological, biomolecular, and nutrient levels when cultivated in the presence of nano or bulk ZnO or CuO or Zn_2^+ or Cu₂⁺ ions. Journal of Hazardous Materials 419:126493. [doi:](http://dx.doi.org/10.1016/j.jhazmat.2021.126493) [10.1016/j.jhazmat.2021.126493.](http://dx.doi.org/10.1016/j.jhazmat.2021.126493)
- Ajouri A, Asgedom H, Becker M. 2004. Seed priming enhances germination and seedling growth of barley under conditions of P and Zn deficiency. Journal of Plant Nutrition and Soil Science 167:630–636. [doi: 10.1002/jpln.200420425.](http://dx.doi.org/10.1002/jpln.200420425)
- Amini R. 2013. Drought stress tolerance of barley (*Hordeum vulgare* L.) affected by priming with PEG. International Journal of Farming and Allied Sciences 2:803–808.
- Amooaghaie R, Nikzad K. 2013. The role of nitric oxide in priming-induced low-temperature tolerance in two genotypes of tomato. Seed Science Research 23:123-131. [doi:](http://dx.doi.org/10.1017/s0960258513000068) [10.1017/s0960258513000068.](http://dx.doi.org/10.1017/s0960258513000068)
- Anwar MP, Ahmed MK, Islam AKMM, Hossain MD, Uddin FMJ. 2020. Improvement of weed competitiveness and yield performance of dry direct seeded rice through seed priming. Turkish Journal of Weed Science 23:15–23.
- Anwar MP, Jahan R, Rahman MR, Islam AKMM, Uddin FMJ. 2021. Seed priming for increased seed germination and enhanced seedling vigor of winter rice. IOP Conference Series: Earth and Environmental Science 756:012047. [doi:](http://dx.doi.org/10.1088/1755-1315/756/1/012047) [10.1088/1755-1315/756/1/012047.](http://dx.doi.org/10.1088/1755-1315/756/1/012047)
- Arief R, Koes F, Komalasari O. 2020. Effects of seed storage duration and matriconditioning materials on germination and seedling characteristics of maize. AGRIVITA Journal of Agricultural Science 42:425. [doi: 10.17503/agrivita.v42i3.2034.](http://dx.doi.org/10.17503/agrivita.v42i3.2034)
- Asaduzzaman M, Huqe M, Uddin M, Hossain M, Haque M. 2021. Seed priming improves germination and early seedling growth in wheat under control and drought condition. Journal of Bangladesh Agricultural University 19:184–191. [doi: 10.5455/jbau.73529.](http://dx.doi.org/10.5455/jbau.73529)
- Ashraf M, Foolad M. 2005. Pre-sowing seed treatment—a shotgun approach to improve germination, plant growth, and crop yield under saline and non-saline conditions. In: Advances in Agronomy. Elsevier. p. 223–271. [doi:](http://dx.doi.org/10.1016/s0065-2113(05)88006-x) [10.1016/s0065-2113\(05\)88006-x.](http://dx.doi.org/10.1016/s0065-2113(05)88006-x)
- Asseng S, Ewert F, Martre P, Rotter RP, Zhu Y. 2014. Rising temperatures reduce global wheat production. Nature Climate Change 5:143–147. [doi:](http://dx.doi.org/10.1038/nclimate2470) [10.1038/nclimate2470.](http://dx.doi.org/10.1038/nclimate2470)
- Aydinoğlu B, Shabani A, Safavi SM. 2019. Impact of priming on seed germination, seedling growth and gene expression in common vetch under salinity stress. Cellular and Molecular Biology 65:18–24. [doi: 10.14715/cmb/2019.65.3.3.](http://dx.doi.org/10.14715/cmb/2019.65.3.3)
- Baloch MJ, Dunwell J, Khakwani AA, Dennett M, Jatoi WA, Channa SA. 2012. Assessment of wheat cultivars for drought tolerance via osmotic stress imposed at early seedling growth stages. Journal of Agricultural Research 50:299–310.
- Baque A, Nahar M, Yeasmin M, Quamruzzaman M, Rahman A, Azad MJ, Biswas PK. 2016. Germination behavior of wheat (*Triticum Aestivum* L.) as influenced by polyethylene glycol (PEG). Universal Journal of Agricultural Research 4:86–91. [doi: 10.13189/ujar.2016.040304.](http://dx.doi.org/10.13189/ujar.2016.040304)
- BARI. 2007. Annual Report 2017–2018. Bangladesh Agricultural Research Institute. Gazipur, Dhaka, Bangladesh.
- Barma NCD, Hossain A, Hakim MA, Mottaleb KA, Alam MA, Reza MMA, Rohman MM. 2019. Progress and challenges of wheat production in the era of climate change: A bangladesh perspective. In: Wheat Production in Changing Environments. Springer Singapore. p. 615–679. [doi: 10.1007/978-981-13-6883-7_24.](http://dx.doi.org/10.1007/978-981-13-6883-7_24)
- BBS. 2020. Statistical Year Book of Bangladesh. Bangladesh Bureau of Statistics, Government of the People?s Republic of Bangladesh, Dhaka, Bangladesh.
- Bose B, Kumar M, Singhal RK, Mondal S. 2018. Impact of seed priming on the modulation of physico-chemical and molecular processes during germination, growth, and development of crops. In: Advances in Seed Priming. Springer Singapore. p. 23–40. [doi: 10.1007/978-981-13-](http://dx.doi.org/10.1007/978-981-13-0032-5_2) [0032-5_2.](http://dx.doi.org/10.1007/978-981-13-0032-5_2)
- Braun HJ, Atlin G, Payne T. 2010. Multi-location testing as a tool to identify plant response to global climate change. In: Climate change and crop production. CABI. p. 115–138. [doi:](http://dx.doi.org/10.1079/9781845936334.0115) [10.1079/9781845936334.0115.](http://dx.doi.org/10.1079/9781845936334.0115)
- Chakraborty P, Dwivedi P. 2021. Seed priming and its role in mitigating heat stress responses in crop plants. Journal of Soil Science and Plant Nutrition 21:1718–1734. [doi: 10.1007/s42729-](http://dx.doi.org/10.1007/s42729-021-00474-4) [021-00474-4.](http://dx.doi.org/10.1007/s42729-021-00474-4)
- Chen X, Zhang R, Xing Y, Jiang B, Li B, Xu X, Zhou Y. 2021. The efficacy of different seed priming agents for promoting sorghum germination under salt stress. PLOS ONE 16:e0245505. [doi:](http://dx.doi.org/10.1371/journal.pone.0245505) [10.1371/journal.pone.0245505.](http://dx.doi.org/10.1371/journal.pone.0245505)
- Cherel I. 2004. Regulation of K^+ channel activities in plants: from physiological to molecular aspects. Journal of Experimental Botany 55:337–351. [doi:](http://dx.doi.org/10.1093/jxb/erh028) [10.1093/jxb/erh028.](http://dx.doi.org/10.1093/jxb/erh028)
- Chunthaburee S, Sanitchon J, Pattanagul W, Theerakulpisut P. 2014. Alleviation of salt stress in seedlings of black glutinous rice by seed priming with spermidine and gibberellic acid. Notulae Botanicae Horti Agrobotanici Cluj-Napoca 42:405–413. [doi: 10.15835/nbha4229688.](http://dx.doi.org/10.15835/nbha4229688)
- da Cruz RP, Milach SCK. 2004. Cold tolerance at the germination stage of rice: methods of evaluation and characterization of genotypes. Scientia Agricola 61:1–8. [doi: 10.1590/s0103-](http://dx.doi.org/10.1590/s0103-90162004000100001) [90162004000100001.](http://dx.doi.org/10.1590/s0103-90162004000100001)
- FAOSTAT. 2019. Food and Agriculture Organization of the United Nations. Rome, Italy.
- FAOSTAT. 2021. Food and Agriculture Organization of the United Nations. Rome, Italy.
- Farooq M, Basra SMA, Ahmad N. 2007a. Improving the performance of transplanted rice by seed priming. Plant Growth Regulation 51:129–137. [doi: 10.1007/s10725-006-9155-x.](http://dx.doi.org/10.1007/s10725-006-9155-x)
- Farooq M, Basra SMA, Khan MB. 2007b. Seed priming improves growth of nursery seedlings and yield of transplanted rice. Archives of Agronomy and Soil Science 53:315–326. [doi:](http://dx.doi.org/10.1080/03650340701226166) [10.1080/03650340701226166.](http://dx.doi.org/10.1080/03650340701226166)
- Farooq M, Basra SMA, Rehman H, Saleem BA. 2008. Seed priming enhances the performance of late sown wheat (*Triticum aestivum* L.) by improving chilling tolerance. Journal of Agronomy and Crop Science 194:55–60. [doi: 10.1111/j.1439-](http://dx.doi.org/10.1111/j.1439-037x.2007.00287.x) [037x.2007.00287.x.](http://dx.doi.org/10.1111/j.1439-037x.2007.00287.x)
- Farooq M, Basra SMA, Tabassum R, Afzal I. 2006. Enhancing the performance of direct seeded fine rice by seed priming. Plant Production Science 9:446–456. [doi: 10.1626/pps.9.446.](http://dx.doi.org/10.1626/pps.9.446)
- Farooq M, Hussain M, Habib MM, Khan MS, Ahmad I, Farooq S, Siddique KHM. 2020a. Influence of seed priming techniques on grain yield and economic returns of bread wheat planted at different spacings. Crop and Pasture Science 71:725. [doi: 10.1071/cp20065.](http://dx.doi.org/10.1071/cp20065)
- Farooq M, Rehman A, Al-Alawi AK, Al-Busaidi WM, Lee DJ. 2020b. Integrated use of seed priming and biochar improves salt tolerance in cowpea. Scientia Horticulturae 272:109507. [doi:](http://dx.doi.org/10.1016/j.scienta.2020.109507) [10.1016/j.scienta.2020.109507.](http://dx.doi.org/10.1016/j.scienta.2020.109507)
- Gahtyari NC, Jaiswal JP, Talha M, Choudhary R, Uniyal M, Kumar N. 2017. Effect of osmotic

stress and seed priming on wheat seed germination traits. International Journal of Current Microbiology and Applied Sciences 6:2799–2809. [doi: 10.20546/ijcmas.2017.604.323.](http://dx.doi.org/10.20546/ijcmas.2017.604.323)

- Gobinathan P, Murali PV, Panneerselvam R. 2009. Interactive effects of calcium chloride on salinityinduced proline metabolism in pennisetum typoidies. Advances in Biological Research 3:168– 173.
- Gomez K, Gomez A. 1984. Statistical Procedure for Agricultural Research. John Wiely and Sons, New York, USA.
- Hasan MN, Salam MA, Chowdhury MMI, Sultana M, Islam N. 2016. Effect of osmopriming on germination of rice seed. Bangladesh Journal of Agricultural Research 41:451–460. [doi:](http://dx.doi.org/10.3329/bjar.v41i3.29717) [10.3329/bjar.v41i3.29717.](http://dx.doi.org/10.3329/bjar.v41i3.29717)
- Hussain HA, Hussain S, Khaliq A, Ashraf U, Anjum SA, Men S, Wang L. 2018. Chilling and drought stresses in crop plants: Implications, cross talk, and potential management opportunities. Frontiers in Plant Science 9:393. [doi:](http://dx.doi.org/10.3389/fpls.2018.00393) [10.3389/fpls.2018.00393.](http://dx.doi.org/10.3389/fpls.2018.00393)
- Hussain S, Hussain S, Khaliq A, Ali S, Khan I. 2019. Physiological, biochemical, and molecular aspects of seed priming. In: Priming and Pretreatment of Seeds and Seedlings. Springer Singapore. p. 43–62. [doi: 10.1007/978-981-13-8625-](http://dx.doi.org/10.1007/978-981-13-8625-1_3) [1_3.](http://dx.doi.org/10.1007/978-981-13-8625-1_3)
- Hussain S, Jamil M, Napar AA, Rahman R, Bano A, Afzal F, Kazi AG, Mujeeb-Kazi A. 2015a. Heat stress in wheat and interdisciplinary approaches for yield maximization. In: Plant-Environment Interaction. John Wiley & Sons, Ltd. p. 161–183. [doi: 10.1002/9781119081005.ch9.](http://dx.doi.org/10.1002/9781119081005.ch9)
- Hussain S, Zheng M, Khan F, Khaliq A, Fahad S, Peng S, Huang J, Cui K, Nie L. 2015b. Benefits of rice seed priming are offset permanently by prolonged storage and the storage conditions. Scientific Reports 5. [doi: 10.1038/srep08101.](http://dx.doi.org/10.1038/srep08101)
- Ibrahim EAA. 2019. Fundamental processes involved in seed priming. In: Priming and Pretreatment of Seeds and Seedlings. Springer Singapore. p. 63–115. [doi: 10.1007/978-981-13-8625-1_4.](http://dx.doi.org/10.1007/978-981-13-8625-1_4)
- Iqbal S. 2020. Influence of seed priming with $CuSO₄$ and $ZnSO_4$ on germination and seedling growth of oat under NaCl stress. Pure and Applied Biology 9:897–912. [doi: 10.19045/bspab.2020.90094.](http://dx.doi.org/10.19045/bspab.2020.90094)
- Islam MS. 2021. Growth and yield performance of selected wheat genotypes at variable irrigation management. Journal of Advanced Agriculture & Horticulture Research 1:24–32.
- Islam R, Mukherjee A, Hossin M. 2012. Effect of osmopriming on rice seed germination and seedling growth. Journal of the Bangladesh Agricultural University 10:15–20. [doi: 10.3329/jbau.v10i1.12013.](http://dx.doi.org/10.3329/jbau.v10i1.12013)
- Jisha KC, Vijayakumari K, Puthur JT. 2012. Seed priming for abiotic stress tolerance: an overview. Acta Physiologiae Plantarum 35:1381–1396. [doi:](http://dx.doi.org/10.1007/s11738-012-1186-5) [10.1007/s11738-012-1186-5.](http://dx.doi.org/10.1007/s11738-012-1186-5)
- Joshi A, Kaur S, Dharamvir K, Nayyar H, Verma G. 2018. Multi-walled carbon nanotubes applied through seed-priming influence early germination, root hair, growth and yield of bread wheat (*Triticum aestivum* L.). Journal of the Science of Food and Agriculture [doi: 10.1002/jsfa.8818.](http://dx.doi.org/10.1002/jsfa.8818)
- Kaczmarek M, Fedorowicz-Strońska O, Głowacka K, Waśkiewicz A, Sadowski J. 2016. CaCl2 treatment improves drought stress tolerance in barley (*Hordeum vulgare* L.). Acta Physiologiae Plantarum 39:41. [doi: 10.1007/s11738-016-2336-y.](http://dx.doi.org/10.1007/s11738-016-2336-y)
- Kamrul MK, Hore R. 2021. Forecasting of rainfall and temperature based on the analysis of historical data and future impacts. Web of Scientist: International Scientific Research Journal 2:353–372.
- Karim MN, Sani MNH, Uddain J, Azad MOK, Kabir MS, Rahman MS, Choi KY, Naznin MT. 2020. Stimulatory effect of seed priming as pretreatment factors on germination and yield performance of yard long bean (*Vigna unguiculata*). Horticulturae 6:104. [doi: 10.3390/horti](http://dx.doi.org/10.3390/horticulturae6040104)[culturae6040104.](http://dx.doi.org/10.3390/horticulturae6040104)
- Khan A, Shafi M, Bakht J, Anwar S, Khan MO. 2020. Effect of salinity (NaCl) and seed priming $(CaCl₂)$ on biochemical parameters and biological yield of wheat. Pakistan Journal of Botany 53:779–789. [doi: 10.30848/pjb2021-3\(12\).](http://dx.doi.org/10.30848/pjb2021-3(12))
- Kubala S, Garnczarska M, Wojtyla Ł, Clippe A, Kosmala A, Żmieńko A, Lutts S, Quinet M. 2015. Deciphering priming-induced improvement of rapeseed (*Brassica napus* L.) germination through an integrated transcriptomic and proteomic approach. Plant Science 231:94–113. [doi:](http://dx.doi.org/10.1016/j.plantsci.2014.11.008) [10.1016/j.plantsci.2014.11.008.](http://dx.doi.org/10.1016/j.plantsci.2014.11.008)
- Lee SS, Kim JH. 2000. Total sugars, α-amylase activity, and germination after priming of normal and aged rice seeds. Korean Journal of Crop Science 45:108–111.
- Lutts S, Benincasa P, Wojtyla L, Kubala S, Pace R, Lechowska K, Quinet M, Garnczarska M. 2016. Seed priming: New comprehensive approaches for an old empirical technique. In: New Challenges in Seed Biology - Basic and Translational

Research Driving Seed Technology. InTech. [doi:](http://dx.doi.org/10.5772/64420) [10.5772/64420.](http://dx.doi.org/10.5772/64420)

- Maiti R, Rajkumar D, Jagan M, Pramanik K, Vidyasagar P. 2013. Effect of seed priming on seedling vigour and yield of tomato and chilli. International journal of Bio-resource and Stress Management 4:119–125.
- Mamun AA, Naher UA, Ali MY. 2018. Effect of seed priming on seed germination and seedling growth of modern rice (*Oryza sativa* L.) varieties. The Agriculturists 16:34–43. [doi:](http://dx.doi.org/10.3329/agric.v16i1.37532) [10.3329/agric.v16i1.37532.](http://dx.doi.org/10.3329/agric.v16i1.37532)
- Mondal S, Vijai P, Bose B. 2011. Role of seed hardening in rice variety swarna (MTU 7029). Research Journal of Seed Science 4:157–165. [doi:](http://dx.doi.org/10.3923/rjss.2011.157.165) [10.3923/rjss.2011.157.165.](http://dx.doi.org/10.3923/rjss.2011.157.165)
- Mouradi M, Bouizgaren A, Farissi M, Makoudi B, Kabbadj A, Very AA, Sentenac H, Qaddoury A, Ghoulam C. 2016. Osmopriming improves seeds germination, growth, antioxidant responses and membrane stability during early stage of moroccan alfalfa populations under water deficit. Chilean journal of agricultural research 76:265– 272. [doi: 10.4067/s0718-58392016000300002.](http://dx.doi.org/10.4067/s0718-58392016000300002)
- Natasha K. 2021. Comparative Effect of sodium chloride, potassium chloride and combined salt stress on germination and growth of *Triticum aestivum* L. (Var. Atta Habib). Pure and Applied Biology 10:1450–1465. [doi: 10.19045/bspab.2021-](http://dx.doi.org/10.19045/bspab.2021-100151) [100151.](http://dx.doi.org/10.19045/bspab.2021-100151)
- Nawaz J, Hussain M, Jabbar A, Nadeem GA, Sajid M, Subtain MU, Shabbir I. 2013. Seed priming a technique. International Journal of Agriculture and Crop Sciences 6:1373.
- Pant B, Bose B. 2016. Mitigation of the influence of PEG-6000 imposed water stress on germination of halo primed rice seeds. International Journal of Agriculture, Environment and Biotechnology 9:275. [doi: 10.5958/2230-732x.2016.00036.x.](http://dx.doi.org/10.5958/2230-732x.2016.00036.x)
- Paparella S, Araújo SS, Rossi G, Wijayasinghe M, Carbonera D, Balestrazzi A. 2015. Seed priming: state of the art and new perspectives. Plant Cell Reports 34:1281–1293. [doi: 10.1007/s00299-015-](http://dx.doi.org/10.1007/s00299-015-1784-y) [1784-y.](http://dx.doi.org/10.1007/s00299-015-1784-y)
- Patade VY, Bhargava S, Suprasanna P. 2009. Halopriming imparts tolerance to salt and PEG induced drought stress in sugarcane. Agriculture, Ecosystems & Environment 134:24–28. [doi:](http://dx.doi.org/10.1016/j.agee.2009.07.003) [10.1016/j.agee.2009.07.003.](http://dx.doi.org/10.1016/j.agee.2009.07.003)
- Patanè C, Cavallaro V, Cosentino SL. 2009. Germination and radicle growth in unprimed and primed seeds of sweet sorghum as affected by reduced

water potential in NaCl at different temperatures. Industrial Crops and Products 30:1–8. [doi:](http://dx.doi.org/10.1016/j.indcrop.2008.12.005) [10.1016/j.indcrop.2008.12.005.](http://dx.doi.org/10.1016/j.indcrop.2008.12.005)

- Prażak R, Święciło A, Krzepiłko A, Michałek S, Arczewska M. 2020. Impact of ag nanoparticles on seed germination and seedling growth of green beans in normal and chill temperatures. Agriculture 10:312. [doi: 10.3390/agriculture10080312.](http://dx.doi.org/10.3390/agriculture10080312)
- Rahman S, Hasan MK. 2009. Wheat in Bangladesh: Yield growth, production performance and determinants. Soybean and Wheat Crops: Growth, Fertilization and Yield. Nova Science Publishers, New York.
- Rai-Kalal P, Jajoo A. 2021. Priming with zinc oxide nanoparticles improve germination and photosynthetic performance in wheat. Plant Physiology and Biochemistry 160:341–351. [doi:](http://dx.doi.org/10.1016/j.plaphy.2021.01.032) [10.1016/j.plaphy.2021.01.032.](http://dx.doi.org/10.1016/j.plaphy.2021.01.032)
- Salah SM, Yajing G, Dongdong C, Jie L, Aamir N, Qijuan H, Weimin H, Mingyu N, Jin H. 2015. Seed priming with polyethylene glycol regulating the physiological and molecular mechanism in rice (*Oryza sativa* L.) under nano-ZnO stress. Scientific Reports 5. [doi: 10.1038/srep14278.](http://dx.doi.org/10.1038/srep14278)
- Salehzade H, Izadkhah Shishvan M, Chiyasi M. 2009. Effect of seed priming on germination and seedling growth of wheat (*Triticum aestivum* L.). Journal of Biological Science 4:629–631.
- Shewry PR, Hawkesford MJ, Piironen V, Lampi AM, Gebruers K, Boros D, Andersson AAM, Åman P, Rakszegi M, Bedo Z, Ward JL. 2013. Natural variation in grain composition of wheat and related cereals. Journal of Agricultural and Food Chemistry 61:8295–8303. [doi: 10.1021/jf3054092.](http://dx.doi.org/10.1021/jf3054092)
- Sisodia A, Padhi M, Pal AK, Barman K, Singh AK. 2018. Seed priming on germination, growth and flowering in flowers and ornamental trees. In: Advances in Seed Priming. Springer Singapore. p. 263–288. [doi: 10.1007/978-981-13-0032-5_14.](http://dx.doi.org/10.1007/978-981-13-0032-5_14)
- Srivastava AK, Bose B. 2012. Effect of nitrate seed priming on phenology, growth rate and yield attributes in rice (*Oryza sativa* L.). Vegetos 25:174– 181.
- Sun YY, Sun YJ, Wang MT, LI XY, Guo X, Hu R, Ma J. 2010. Effects of seed priming on germination and seedling growth under water stress in rice. Acta Agronomica Sinica 36:1931–1940. [doi:](http://dx.doi.org/10.1016/s1875-2780(09)60085-7) [10.1016/s1875-2780\(09\)60085-7.](http://dx.doi.org/10.1016/s1875-2780(09)60085-7)
- Thor K. 2019. Calcium—nutrient and messenger. Frontiers in Plant Science 10. [doi:](http://dx.doi.org/10.3389/fpls.2019.00440) [10.3389/fpls.2019.00440.](http://dx.doi.org/10.3389/fpls.2019.00440)
- Varier A, Vari AK, Dadlani M. 2010. The subcellular basis of seed priming. Current Science :450–456.
- Wahid A, Noreen A, Basra SMA, Gelani S, Farooq M. 2008. Priming-induced metabolic changes in sunflower (*Helianthus annuus*) achenes improve germination and seedling growth. Botanical Studies 49:343–350.
- Wei W, Li QT, Chu YN, Reiter RJ, Yu XM, Zhu DH, Zhang WK, Ma B, Lin Q, Zhang JS, Chen SY. 2014. Melatonin enhances plant growth and abiotic stress tolerance in soybean plants. Journal of Experimental Botany 66:695–707. [doi:](http://dx.doi.org/10.1093/jxb/eru392) [10.1093/jxb/eru392.](http://dx.doi.org/10.1093/jxb/eru392)
- Zampieri M, Ceglar A, Dentener F, Toreti A. 2017. Wheat yield loss attributable to heat waves, drought and water excess at the global, national and subnational scales. Environmental Research Letters 12:064008. [doi: 10.1088/1748-](http://dx.doi.org/10.1088/1748-9326/aa723b) [9326/aa723b.](http://dx.doi.org/10.1088/1748-9326/aa723b)
- Zhang F, Yu J, Johnston CR, Wang Y, Zhu K, Lu F, Zhang Z, Zou J. 2015. Seed priming with polyethylene glycol induces physiological changes in sorghum (*Sorghum bicolor* L. Moench) seedlings under suboptimal soil moisture environments. PLOS ONE 10:e0140620. [doi:](http://dx.doi.org/10.1371/journal.pone.0140620) [10.1371/journal.pone.0140620.](http://dx.doi.org/10.1371/journal.pone.0140620)
- Zheng M, Tao Y, Hussain S, Jiang Q, Peng S, Huang J, Cui K, Nie L. 2015. Seed priming in dry direct-seeded rice: consequences for emergence, seedling growth and associated metabolic events under drought stress. Plant Growth Regulation 78:167–178. [doi: 10.1007/s10725-015-0083-5.](http://dx.doi.org/10.1007/s10725-015-0083-5)

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The Official Journal of the **Farm to Fork Foundation** ISSN: 2518–2021 (print) ISSN: 2415–4474 (electronic) <http://www.f2ffoundation.org/faa>